Steady-Glo® Luciferase Assay System

INSTRUCTIONS FOR USE OF PRODUCTS E2510, E2520 AND E2550.

Quick PROTOCOL

Standard Protocol for the Steady-Glo® Assay System

Steady-Glo® Luciferase Assay System yields reliable and robust results in high-throughput screening applications. The Steady-Glo® Reagent provides stable luminescence with a half-life of greater than 5 hours when used with common cell culture media.

1. Remove 96- or 384-well plates containing mammalian cells from the incubator. The plates used must be compatible with the luminometer being used. For best results, equilibrate cultured cells to room temperature before performing Step 2.
2. To each plate well, add a volume of Steady-Glo® Reagent equal to the volume of culture medium in the well, and mix. (For 96-well plates, typically 100µl of reagent is added to cells grown in 100µl of medium. For 384-well plates, typically 30µl of reagent is added to cells grown in 30µl of medium.)
3. Wait at least 5 minutes to allow cell lysis, then measure luminescence in a luminometer.

See additional protocol information in Technical Manual #TM051, available online at: **www.promega.com**

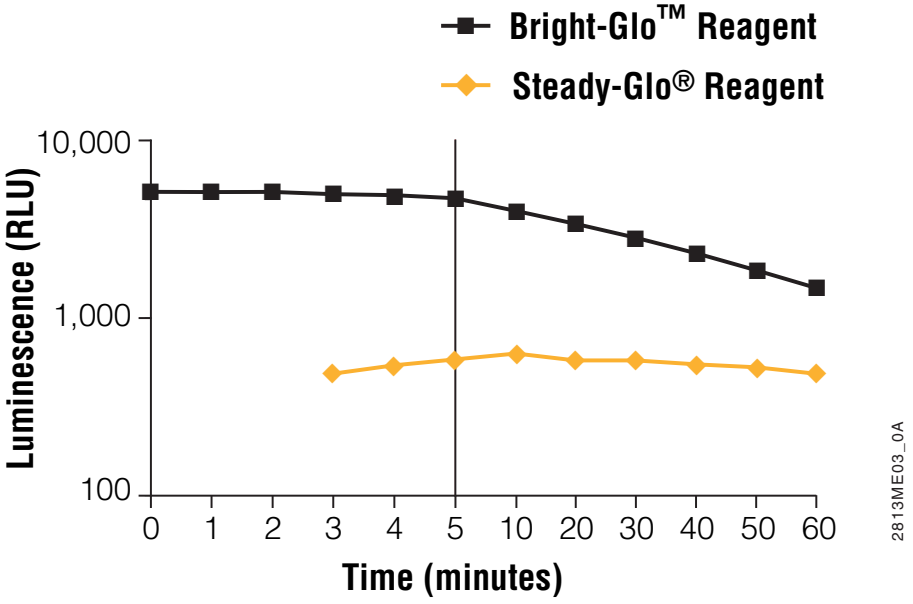
Characteristics of Steady-Glo® Assay Reagent.

	Steady-Glo® Reagent
Format	NH or H
Process	Batch
Sensitivity	Lower light output
Signal Half-Life	≥5 hours
Change in Luminescence Per 96-Well Plate*	<1%
Cell Lysis Time	~5 minutes

NH = Nonhomogeneous assay format (when used with Glo Lysis Buffer, Cat.# E2661).

H = Homogeneous assay format.

*Percent change in luminescence was determined as follows: Reagent was added to all wells, the plate was read, and the signals from wells 1 and 96 were compared.



ORDERING/TECHNICAL INFORMATION:
www.promega.com • Phone 608-274-4330 or 800-356-9526 • Fax 608-277-2601

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Part #9FB037

Bright-Glo™ Luciferase Assay System

INSTRUCTIONS FOR USE OF PRODUCTS E2610, E2620 AND E2650.

Quick PROTOCOL

Standard Protocol for the Bright-Glo™ Assay System

Bright-Glo™ Reagent provides four- to tenfold more light output than other extended half-life luciferase reagents, depending on the cell line and medium used. Signal half-life is approximately 30 minutes in commonly used cell culture media, which is sufficient to read a 96-well plate with less than 5% signal decay.

1. Remove 96- or 384-well plates containing mammalian cells from the incubator. The plates used must be compatible with the luminometer being used. For best results, equilibrate cultured cells to room temperature before performing Step 2.
2. To each well add a volume of Bright-Glo™ Reagent equal to the volume of culture medium in the well, and mix. (For 96-well plates, typically 100µl of reagent is added to cells in 100µl of culture medium. For 384-well plates, typically 30µl of reagent is added to cells grown in 30µl of medium.)
3. Wait at least 2 minutes to allow cell lysis, then measure luminescence in a luminometer.

See additional protocol information in Technical Manual #TM052, available online at: ***www.promega.com***

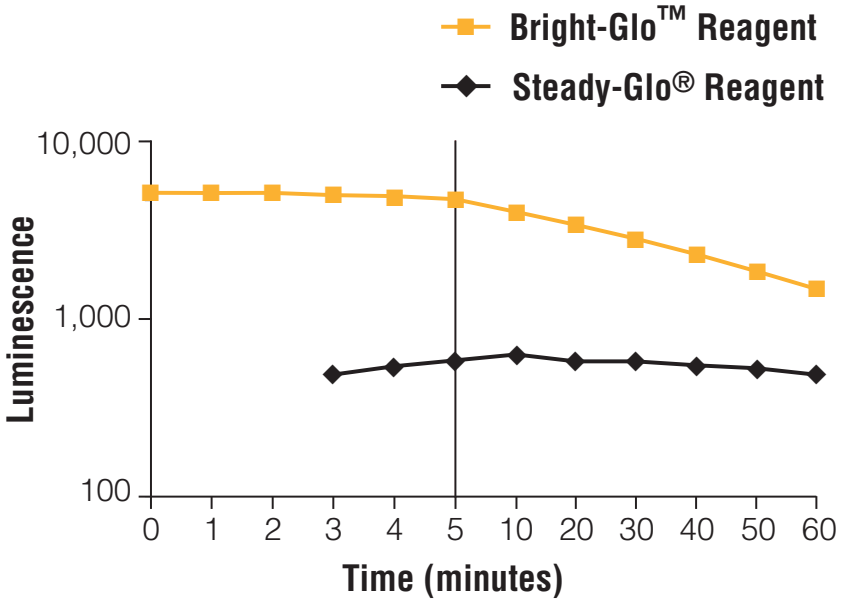
Characteristics of Bright-Glo™ Assay Reagent.

	Bright-Glo™ Reagent
Format	NH or H
Process	Continuous
Sensitivity	Maximum light output
Signal Half-Life	~ 30 minutes
Change in Luminescence Per 96-Well Plate*	<5%
Cell Lysis Time	~ 2 minutes

NH = Nonhomogeneous assay format (when used with Glo Lysis Buffer, Cat.# E2661).

H = Homogeneous assay format.

*Percent change in luminescence was determined as follows: Reagent was added to all wells, the plate was read and the signals from wells 1 and 96 were compared.



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